

Phosphorus deficiency enhances water deficit impact on some morphological and physiological traits in four faba bean (*Vicia faba* L.) varieties

Khawla Oukaltouma,^{1,2} Ahmed El Moukhtari,¹ Yahya Lahrizi,¹ Mohammed Mouradi,³ Mohamed Farissi,³ Anne Willems,⁴ Ahmed Qaddoury,¹ Faouzi Bekkaoui,^{2,5} Cherki Ghoulam^{1,2}

¹Team of Biotechnology and Symbiosis Agrophysiology, Faculty of Sciences and Techniques, UCA, Marrakech, Morocco; ²Agrobiosciences Program, Mohamed VI Polytechnic University, Benguerir, Morocco; ³Laboratory of Biotechnology and Sustainable Development of Natural Resources (B2DRN), Polydisciplinary Faculty, USMS, Beni Mellal, Morocco; ⁴Laboratory of Microbiology, Faculty of Sciences, Ghent University, Ghent, Belgium; ⁵National Institute of Agronomic Research (INRA), Rabat, Morocco

Highlights

- Varieties tolerant to low phosphorus and water deficiency are needed in arid and semi-arid regions with low P.
- Phosphorus nutrition is advantageous for mitigating the effect of water deficit on faba bean (*Vicia faba*) plants and conversely, deficient P supply reduces resilience to water deficit.
- The faba bean variety Aguadulce showed high tolerance to the combined effect of water deficit and phosphorus deficiency in terms of growth, leaf water potential, stomatal conductance, membrane permeability and glycine betaine accumulation.
- Reina Mora was the least tolerant variety to water deficit combined with P limitation.
- Combined limitation of water and phosphorus induced accumulation of glycine betaine in leaves.

Abstract

Moroccan soils, generally present low available phosphorus (P) levels which occur in almost all arid and semi-arid regions. Faba bean is one of the most significant crops in Morocco and is

influenced by these constraints that affect its nutrient uptake and nitrogen fixation capacity and hence plants development. Therefore, we evaluated the response of four *Vicia faba* varieties - Aguadulce (Ag), Alfia (Al), Luz de Otono (LO) and Reina Mora (RM) -, grown under biological nitrogen fixation to water deficit and two phosphorus levels. The trial was conducted under greenhouse conditions and water stress was induced by keeping pots at 40% substrate field capacity (FC) versus 80% FC for the controls while phosphorus deficiency treatment consisted in the application of 25 $\mu\text{mol P plant}^{-1} \text{ week}^{-1}$ versus 125 $\mu\text{mol P plant}^{-1} \text{ week}^{-1}$ for sufficient P treatment. The results revealed a significant effect of water deficit and phosphorus deficiency either alone or combined on plants dry weights, leaf water parameters and nutrient concentrations. However sufficient phosphorus supply mitigated the adverse effects of water deficit on faba bean. We noticed significant differences between the studied varieties. Ag showed high performance concerning dry weights (1.25 g and 1.88 g plant^{-1} respectively for shoot and root) and high concentration of nitrogen N (4.7%) and P (0.27 mg g^{-1} DW) and was then qualified as the most tolerant variety to water deficit combined with P limitation. While RM was the least tolerant variety, as it showed the lowest dry weights (0.51 g and 1.4 g plant^{-1} respectively for shoot and root) and concentration (2.74% and 0.19 mg g^{-1} DW respectively for N and P). The tolerance was related to the ability to ensure efficient osmoregulation by glycine betaine accumulation, to keep leaf water balance and cell membrane stability that contribute together with adequate symbiotic nitrogen fixation to plant growth performance under combined stresses.

Correspondence: Cherki Ghoulam, Team of Biotechnology and Symbiosis Agrophysiology, Faculty of Sciences and Techniques, UCA, Marrakech, Morocco.

E-mail: c.ghoulam@uca.ma

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Introduction

Faba bean (*Vicia faba* L.) cultivated land worldwide was 3.7 million ha by 1980 and declined to 2.1 million ha by 2014 (FAO, 2017). The inclusion of faba bean in human and animal diets has

health benefits because it presents a high protein content and is a source of several nutrients including Fe, Mg, Zn, K and Ca, amino acids, carbohydrates, vitamins and essential nutraceuticals (Multari *et al.*, 2015; Koivunen *et al.*, 2016). The addition of pulse crops such as faba beans, in rotation with cereals, improves soil: physical, chemical, and biological properties, soil fertility, disturbs pest and disease cycles and reduces the use of inorganic nitrogen fertilizer through the biological nitrogen fixation (BNF) (Jensen *et al.*, 2012). This capacity of fixation, ranges from 90 to 200 kg N ha⁻¹ (Herridge *et al.*, 2008; Neugschwandtner *et al.*, 2015), with a potential of up to 300 kg N ha⁻¹ (Singh *et al.*, 2013). The nitrogen fixation variations depend on the variety, agronomic practices, soil properties and mainly on the presence of compatible symbiotic rhizobacteria in the soil (Argaw and Mnalku, 2017). However, the BNF is an energy intensive process that requires large amount of phosphorus.

Phosphorus is one of the major plant nutrients limiting growth due to its role in improving the nodulation process and BNF, root and nodule development, plant growth and formation of phosphoglycolate needed in photosynthesis (Kubure *et al.*, 2016). However, approximately 70% of cultivated soils suffer from P limitation (Hinsinger, 2001), making P nutrition a great priority for maintaining good plant production. Only 20% to 30% of applied phosphate is used and absorbed by the plants and the remaining is lost due to interaction with soil components and microorganisms or run-off from soils to lakes, rivers and seas (Kirkby and Johnston, 2008). Therefore, even if soils contain high amounts of phosphorus, it is largely unavailable to the plant owing to its high reactivity with soil components such as iron, aluminium and calcium which result in the formation of highly insoluble forms of phosphorus.

Faba bean plants also require relatively large amounts of water, especially to ensure maximum seed germination and in some cases low water availability could also be a stress to more mature plants. In general, water stress is defined as the reduction of available soil moisture and the increase of plant water loss by transpiration (Jaleel *et al.*, 2009). Climate change has become a threat around the world, causing floods in some areas and water scarcity and drought in others. Morocco is one of the areas that has started to suffer from water limitation especially for rain-fed agro-systems. Moreover, studies of the effect of water stress on plant production reported that water deficit reduced yield by 50% to 82% depending on its severity (Wang *et al.*, 2003; Zlatev and Lidon, 2012). According to Razmjoo *et al.* (2008), plant tolerance to abiotic stresses is complicated by the interaction between various stress factors and the difference in physiological, biochemical and molecular processes involved in plant growth and development. For this reason, the occurrence of water deficit and phosphorus deficiency together represents dual constraints for crop production. Meanwhile, various key changes and adaptation mechanisms related to the tolerance to water deficit or phosphorus limitation, were associated to the decrease of growth and regulation of water status parameters such as biomass, leaf area, stomatal conductance, relative water content (Alghamdi *et al.*, 2015) and the ability of the plants to keep the balance of leaf water potential, the photosynthetic function and membrane stability (Waraich *et al.*, 2011; Kabbadj *et al.*, 2017; Mouradi *et al.*, 2018).

Furthermore, according to Zhu (2002) abiotic stressors such as drought may also induce osmotic and oxidative stress in plants. As a response, plants adopt various mechanisms such as compatible solutes accumulation, antioxidant system induction and reactive oxygen species (ROS) scavenging systems. Compatible solutes are highly soluble compounds that are usually nontoxic at high cellular concentrations (Giri, 2011) and include organic osmolytes like pro-

line and glycine betaine and inorganic osmolytes like K⁺, Ca²⁺ and Na⁺ (Farissi *et al.*, 2013; Bargaz *et al.*, 2015; Kabbadj *et al.*, 2017). With reference to the morpho-physiological adaptation of plants to the combined effect of water deficit and phosphorus limitation, it is documented that their combined effect reduces nutrient uptake, relative water content, leaf water potential and photosynthetic activity (Shubhra *et al.*, 2004; Jemo *et al.*, 2017). However, as most of faba bean metabolism alterations are related to the regulation associated to water status, their change under the combined effect of water deficit and phosphorus limitation will give an understanding of the role of phosphorus level combined with drought on morpho-physiological response of faba bean plants.

The individual effects of water deficit and phosphorus deficiency on Aguadulce, Alfia, Reina Mora and Luz D'Otono varieties as the commonly grown faba bean varieties in Morocco, have been widely studied (Kabbadj *et al.*, 2017; Makoudi *et al.*, 2018; Mouradi *et al.*, 2018) but there is a lack of information on their combined effects on faba bean plants and also of the beneficial effect of sufficient phosphorus fertilization on this species under water deficit. Moreover, an increase in water deficit in the Mediterranean area, as forecast for the coming decades, could be accompanied by a decreased in P soil availability for faba bean. Thus, the identification of physiological and molecular traits that vary in response to the combination of the two stress events remains a challenge. The present research was undertaken to assess the growth and nutrient uptake, as well as physiological and biochemical responses of four faba bean varieties under combined water and phosphorus shortage. Therefore, the aims of this study are to: i) investigate the individual and concurrent effects of drought and P stresses on growth, water status, osmolyte accumulation, nutrient uptake and membrane stability in faba bean; ii) examine the basis of the crop tolerance against drought and P stresses; and iii) assess the performance of four different varieties under drought and/or P stresses. We hypothesize that P shortage makes water deficit more severe and this combination will affect the faba bean metabolism differently compared to a single stress.

Materials and methods

Treatments and trial description

The experiment was set up under greenhouse conditions at the Faculty of Sciences and Techniques of Marrakech, with an average day/night temperature of 25/20°C, an approximate relative humidity of 50 to 80% and a 16 h photoperiod. Four faba bean varieties frequently grown in the Haouz area of Morocco and known for their high nodulation and their different levels of tolerance to water deficit (Kabbaj *et al.*, 2017); Aguadulce (Ag) and Alfia (Al) Moroccan varieties, and Luz d'Otono (LO) and Reina Mora (RM), Spanish ones, were used. The faba bean seeds were surface disinfected by immersion in 5% sodium hypochlorite for 5 min, rinsed four times in sterile distilled water and germinated in sterilized sand for 7 days. Seedlings were transplanted into plastic pots measuring 20 cm deep and 22 cm in diameter filled with 1.5 kg of sterilized sand and peat mixture (4 : 1, v/v). Two seedlings were transplanted per pot. The pot was considered as an experimental unit. Five replicates (ten plants) per treatment per variety were considered. Each transplanted seedling was inoculated with 5 ml *Rhizobium leguminosarum* RhF41 that was isolated and identified from nodules of faba beans grown at the Haouz area, at a rate of 10⁸ colony forming unit (CFU) mL⁻¹.

Plants were irrigated with distilled water and Hoagland nitrogen free solution (100 $\mu\text{mol L}^{-1}$ MgSO_4 , 750 $\mu\text{mol L}^{-1}$ K_2SO_4 , 1650 $\mu\text{mol L}^{-1}$ CaCl_2 , 16 $\mu\text{mol L}^{-1}$ Sequestrene, 6 $\mu\text{mol L}^{-1}$ MnSO_4 , 4 $\mu\text{mol L}^{-1}$ H_3BO_3 , 1 $\mu\text{mol L}^{-1}$ ZnSO_4 , 0.1 $\mu\text{mol L}^{-1}$ NaMoO_4 , 1 $\mu\text{mol L}^{-1}$ CuSO_4); the nutrient solution was applied once a week with no additional nitrogen. Two weeks after transplantation, water was restricted to 40% field capacity (FC) for stressed plants *versus* 80% FC for non-stressed plants (controls) (Kabbadj *et al.*, 2017) while phosphorus deficiency treatment consisted in the application of 25 μmol of P plant^{-1} week^{-1} *versus* 125 μmol of P plant^{-1} week^{-1} for sufficient P treatment, provided as KH_2PO_4 (Makoudi *et al.*, 2018). The combined stress was applied by submitting the plants to water deficit (40% FC) and P deficiency (25 μmol of P plant^{-1} week^{-1}). At the flowering stage; 50 to 55 days after sowing (61 BBCH-scale) and 40 days of stress, leaf water potential, stomatal conductance and photosynthetic performance were measured in situ on the plants under the greenhouse conditions while leaf samples and plants were collected for growth, physiological and biochemical assessment.

Plant dry weight measurement

At flowering stage, plants were harvested and shoots were separated from the roots using sharp scissors. Then, substrate attached to the root was carefully removed using a brush and washed several times using distilled water. Nodules were collected from each plant using metal tweezers. The three plant parts were dried in an oven (Binder™ series FD-S Solid.Line) at 70°C for 48 h and the dry weight was measured for ten plants per treatment per variety.

Three random samples were used for nitrogen, phosphorus, Na^+ , K^+ and Ca^{2+} concentration analyses for each treatment and per variety for the three plant parts, from the ten dried plants.

Leaf area

At the flowering stage, leaf samples were collected and their surfaces were measured using Image J Software (<http://rsb.info.nih.gov/ij/index.html>). Three replications per treatment per variety were used.

Stomatal conductance

The stomatal conductance is a response of ambient CO_2 concentration, leaf air vapor pressure difference and leaf temperature and water status. It was measured on eight plants per treatment per variety, outliers were eliminated and the remaining values were regrouped to form three replications. The measures were taken at noon under $28 \pm 2^\circ\text{C}$ and $58 \pm 4\%$ of relative humidity on leaves with a porometer (SC1 Model, Decagon Devices, version 2012).

Relative water content

Relative water content (RWC) represents the water that a leaf contains relative to its water content at full turgidity. To measure this parameter, fresh weights (FW) of foliar disks of three leaves per treatment and per variety were determined. They were then immersed for 6 h in distilled water to reach full turgidity. After wiping the surface water from leaf disks, their turgid weights (TW) were measured. Then samples were dried for 24 h at 70°C and their dry weights (DW) were determined. The RWC was determined using the following formula (Ghoulam *et al.* 2002).

$$\text{RWC} = \frac{(\text{FW} - \text{DW})}{(\text{TW} - \text{DW})} \times 100 \quad (1)$$

Chlorophyll fluorescence (F_v/F_m)

At 61 BBCH-scale of growth stage, chlorophyll fluorescence which gives an estimation of the degree of photoinhibition (Jifon and Syvertsen, 2003) was measured with a fluorimeter (Handy PEA, Hansatech, England). The illumination of the leaves with a beam of saturating light after their dark adaptation for 20 to 30 min, permits the determination of F_m , F_0 and F_v ($F_m - F_0$) which are respectively the maximal quantum yield of PS II, the minimal fluorescence intensity and the variable fluorescence. The chlorophyll fluorescence of six leaves per treatment per variety was quantified by F_v/F_m ratio and grouped in three replicates.

Leaf water potential

Leaf water potential was measured at noon on six leaf samples belonging to the same foliar rank on stem, per treatment and per variety. Water potential was measured using a pressure chamber (PMS Instrument Co, Model 600, USA).

Membrane permeability

The membrane stability was determined by measuring the electrolyte leakage from leaf disks. Leaf samples were washed 3 times with deionized water and leaf disks were cut and shaken for 24 h in sealed vials containing 10 ml of deionized water. Electrical conductivity of the solution (E1) was determined with a conductivity meter (Hannah Instruments HI8820 N) at 25°C. The foliar disks were then autoclaved for 20 min at 120°C and the electrical conductivity was measured (E2) at 25°C. The following formula was used to express the percentage of electrolyte leakage:

$$\text{Electrolyte leakage (EL)} = \frac{E1}{E2} \times 100 \quad (2)$$

Membrane lipids peroxidation

Lipid peroxidation was assessed according to Šavicka and Skute (2010) by the estimation of malondialdehyde (MDA) accumulation. Leaf samples of 0.5 g were ground in 3 ml of trichloroacetic acid (TCA) 0.1% (w/v). Then, the homogenate was centrifuged at 14000 $\times g$ for 15 min and 1 ml of the obtained supernatant was mixed with 2.5 ml thiobarbituric acid (TBA) 0.5% (w/v) prepared in TCA 20% (w/v). After, incubation at 95°C for 30 min followed by immersion of the reaction tubes in an ice bath and centrifugation at 5000 rpm for 5 min, the absorption of the obtained supernatant was read at 532 nm and 600 nm. The calculation of MDA accumulation of three replications per treatment were done based on the extinction coefficient (ϵ) of 155 $\text{mM}^{-1} \text{cm}^{-1}$.

$$\text{MDA (mM)} = \frac{A532 - A600}{\epsilon} \quad (3)$$

Glycine betaine

The accumulation of osmolytes like glycine betaine (GB) in cells is known to protect organisms against abiotic stresses via osmoregulation or osmoprotection. It was measured using 0.5 g of dried, ground leaves shaken in 20 mL of distilled water for 24 h at 25°C. According to the method of Grieve and Grattan (1983), 0.5 mL of the extract was added to 0.5 mL of H_2SO_4 and was shaken in ice bath for 60 min. A volume of 0.2 mL of Cold KI-I_2 reagent was added to 0.5 mL of the extract and then stored for 16 h at 4°C before centrifugation at 10,000 rpm (8000 $\times g$) for 15 min at 4°C. After elimination of the supernatant, the periodide crystals were dissolved in 9 ml of 1,2-dichloroethane and the absorbance was measured at 365 nm after 2 to 3 h. The concentration was expressed as μmol glycine betaine per g of DW using calibration curves. Three replicates were

performed per treatment and per variety.

P, Na⁺, K⁺ and Ca²⁺ concentrations

For the P, Na⁺, K⁺ and Ca²⁺ concentration measurements, dried samples of 0.5 g (shoot, root, and nodule) were incinerated at 600°C for 6 h in a Thermolyne Tabletop Muffle Furnaces. The ash formed was recovered by adding 3 ml of HCl (10 N). The solution was then filtered and adjusted to 100 mL by distilled water. The resulting solution was used for nutrient analyses (Ghoulam *et al.*, 2002). For P concentration determination, to 1 mL of the prepared solution, 4 ml of distilled water and 5 ml of a mixture of 2.5% (w/v) sodium molybdate and 0.15% (w/v) hydrazine sulphate were added. The resulting solution was heated for 10 min in a 95°C water bath and after colour development the absorbance was measured by spectrometer at 820 nm.

The Na⁺, K⁺ and Ca²⁺ concentrations in the same prepared solution were determined by flame emission photometry (AFP100 Model, Biotech Management Engineering Co. Ltd., UK).

Plant nitrogen concentrations

Shoot and root nitrogen concentrations were determined according to Kjeldahl method. In matrass tubes, 10 mL of concentrated H₂SO₄ were added to 0.5 g dry biomass and 1 g catalyst and then digested for 2 h at 400°C. A distillation unit connected to an Erlenmeyer flask containing a solution of 10 ml of boric acid and 20 ml of NaOH was used. The distillate was recovered and the total nitrogen concentration was determined by titration of 5 ml of the distillate by sulfuric acid (0.01 mol L⁻¹) using bromocresol green and methyl red as colour indicators. Three replicates were performed per treatment and per variety.

Statistical analysis

The experiment was set up using a split-split-plot design with the varieties [Aguadulce (Ag) and Alfia (Al), Luz d'Otono (LO) and Reina Mora (RM)] as the main plot, water regime (40% FC, 80% FC) as the subplot, and P treatments (P0: 25 μmol plant⁻¹ week⁻¹, P1: 125 μmol plant⁻¹ week⁻¹) as the sub-subplot. Each of the four main plots was blocked into five replicates. The data were statistically analysed using IBM SPSS statistics 20 software (IBM Corporation and Others, Armonk, NY, USA) and presented as the mean ± standard error. The means were compared statistically at the level of P<0.05 using Student-Newman-Keuls's multiple-range test (Tables 1-6).

Results

Plant dry biomass

The statistical analysis showed that water deficit and P deficiency either alone or combined, significantly affected (P<0.001, Table 6) shoot, root dry weights. However, no significant difference was observed between the varieties' nodule dry weights in response to the combined water and P deficiency. Moreover, all varieties tested produced markedly higher biomass when water was not limiting, than under deficit conditions regardless of P level. But under the combined stress, the dry biomass reduction was more pronounced for almost all of plant tissues testes and in all of the tested faba bean varieties, compared to their corresponding controls (Figure 1).

Limiting P induced a slight decrease in shoot dry weight which was more pronounced under water deficit with a significant varia-

tion observed for Al, LO and RM varieties. Under the combined stressors, the variety RM presented the lowest biomass of 0.51 g plant⁻¹ versus 1.25 g plant⁻¹ obtained by the Ag variety. The Ag presented the highest biomass and then a less reduction compared to its corresponding control. For root biomass, Ag variety showed the similar reductions under all treatments but produced the highest biomass (1.88 g plant⁻¹) when both stresses were combined, compared to Al variety which produced the lowest biomass (0.87 g plant⁻¹). For nodule dry biomass, water deficit and its combina-

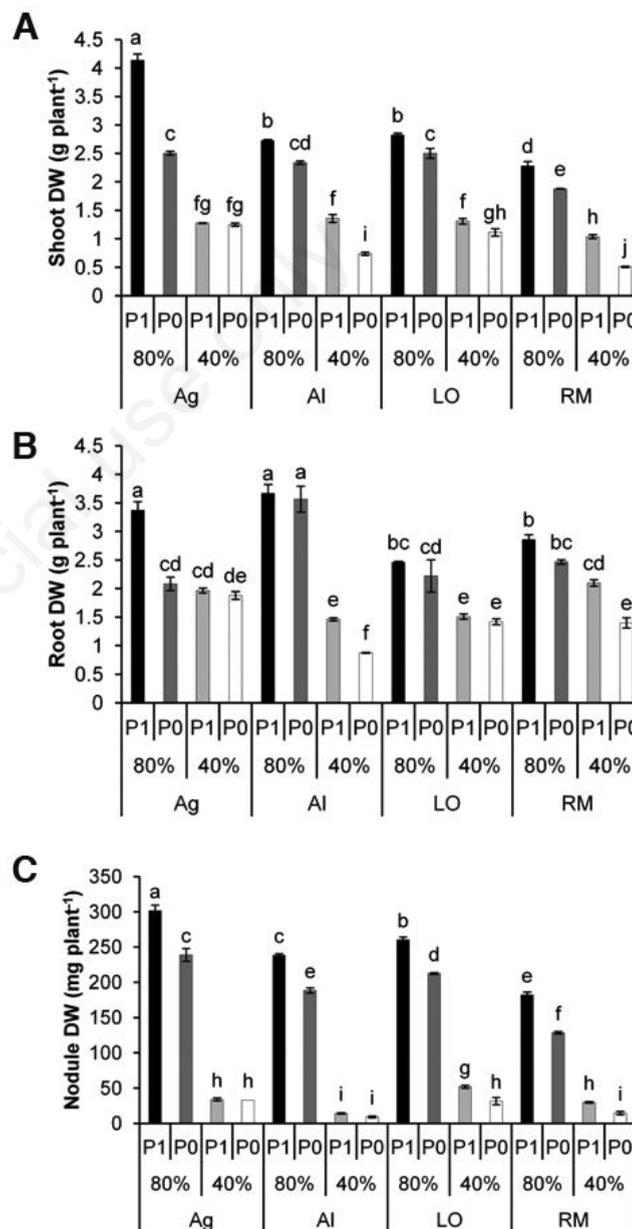


Figure 1. Plants dry biomass (A: Shoot, B: Root, C: Nodule) under water deficit [40% field capacity (FC)] and phosphorus deficiency (25 μM) and their combination (40% FC; 25 μM) in comparison to control condition (80% FC; 125 μM) of four faba bean varieties Ag (Aguadulce), Al (Alfia), LO (Luz de Otono) and RM (Reina Mora). The values represent the means of ten replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

tion with P limitation produced a severe reduction and it was more pronounced in AI and RM varieties, with a reduction rates relative to their corresponding controls of 96% and 92% respectively.

Under combined effect of water deficit and P deficiency, Ag presented the highest shoot, root and nodule dry weight (1.25; 1.88 g plant⁻¹ and 33 mg plant⁻¹ respectively for shoot, root and nodule).

Leaf area

Water stress, P deficiency and their interaction significantly reduced ($P < 0.001$, Table 6) leaf area and no significant difference was observed between the varieties in response to the combined stressors. Under all applied treatments, this parameter varied from 32 to 11 cm² (Figure 2). For sufficient water regime and both P

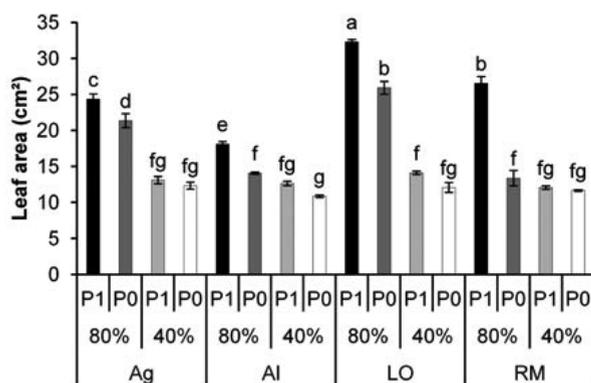


Figure 2. The effect of water deficit [40% field capacity (FC)] and phosphorus deficiency (25 μM) and their combination (40% FC; 25 μM) on four faba bean varieties' Leaf area. The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

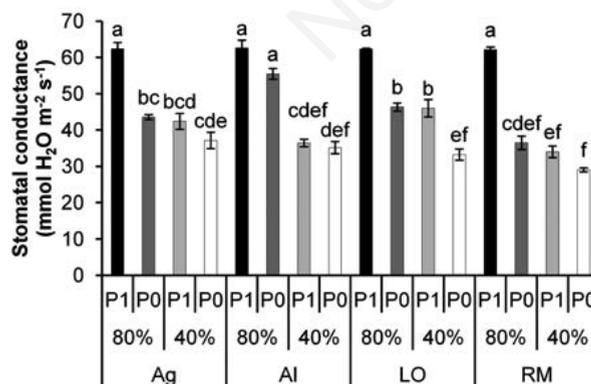


Figure 3. The effect of water deficit [40% field capacity (FC)] and phosphorus deficiency (25 μM) and their combination (40% FC; 25 μM) on stomatal conductance of faba bean varieties. The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

concentrations, the lowest leaf areas were obtained for AI variety and the highest ones for LO variety. However, LO variety showed the highest reduction rate of 62.6% under combined water and P deficiency relative to its control while AI variety showed the lowest reduction rate of 40%.

Stomatal conductance

Under the studied abiotic stresses, water deficit, P limitation and their combination, stomatal conductance decreased significantly ($P < 0.05$, Table 6). In general, this parameter similarly varied for all tested varieties. Under sufficient water and P concentrations, Ag exchanged 62 mmol H₂O m⁻² s⁻¹ (Figure 3). This conductance was reduced under combined water and P deficiency to 37 mmol H₂O m⁻² s⁻¹, representing a decrease of 40%. The highest decrease of stomatal conductance was noticed in the RM variety showing a reduction of 53% under the dual stress compared to its control. The AI and LO varieties showed a reduction of stomatal conductance of 44% and 47% respectively under combined stresses relative to their corresponding controls.

Relative water content

Statistical analysis revealed a significant effect of water deficit and P deficiency separately ($P < 0.001$, Table 6) on the relative water content that leaves could hold in each variety. For control plants, RWC varied from 70% obtained in RM variety to 61.7% in LO variety. Under the combined stresses, RWC varied from 52%, obtained in Ag variety to 43%, obtained in AI variety. The RWC decreased the least in the Ag variety (22%) and the RWC decreased the most in the RM variety 33% relative to their respective controls (Figure 4).

Chlorophyll fluorescence (F_v/F_m)

Results presented in Figure 5 show the responses of four faba bean varieties to water deficit and phosphorus deficiency and their

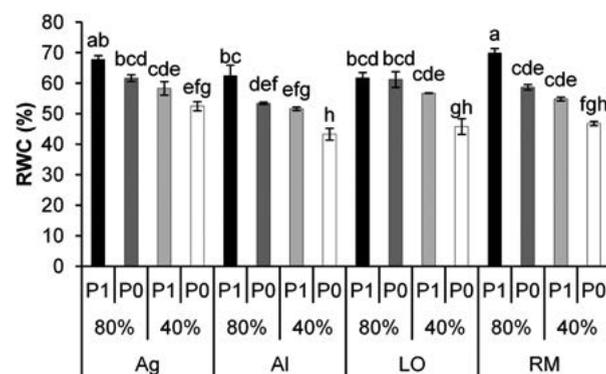


Figure 4. Relative water content (RWC) of 50 days old plants under water deficit [40% field capacity (FC)] and phosphorus deficiency (25 μM) and their combination (40% FC; 25 μM) in comparison to control conditions (80% FC; 125 μM) of four faba bean varieties Ag (Aguadulce), AI (Alfia), LO (Luz de Otono) and RM (Reina Mora). The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

combination regarding PS II activity. Under 80% FC, the maximum quantum yield of PS II varied from 0.833 observed in the variety RM to 0.807 in the LO variety. Under water deficit, this parameter varied from 0.796 to 0.711 in the AI variety in the presence of sufficient P treatment and P deficiency respectively. Therefore, limiting water had a greater effect on efficiency of photosystem II but the effect is decreased with sufficient P. AI variety showed the lowest F_v/F_m value (0.711) under combined stresses and also the greatest reduction rate (14%) relative to its control. Ag variety showed the highest F_v/F_m value (0.765) under combined stresses and also the lowest reduction (7.6%) relative to its control. For statistical analysis, water deficit and phosphorus limitation had a significant individual effect ($P < 0.001$, Table 6) or combined ($P < 0.05$).

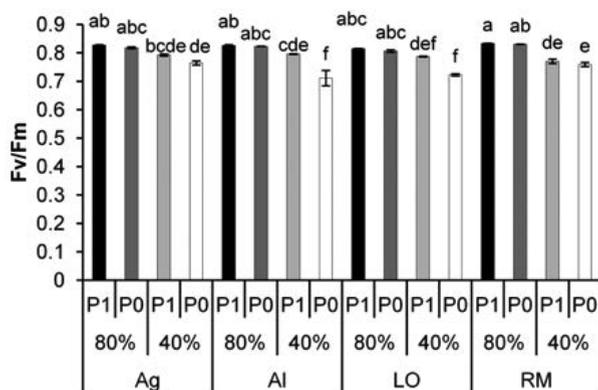


Figure 5. Efficiency of photosystem II (F_v/F_m) under combined water deficit and phosphorus deficiency (40% field capacity; 25 μM). The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

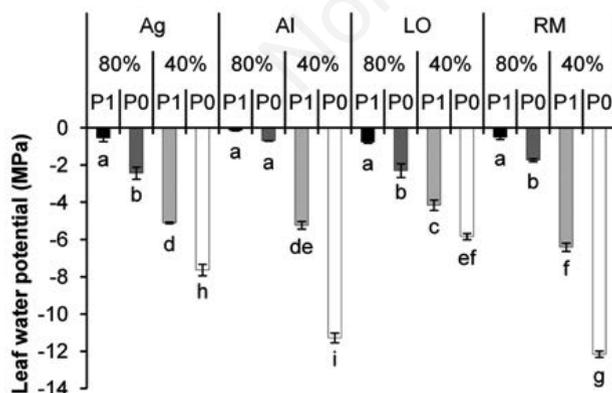


Figure 6. Variation of leaf water potential under water deficit (40% field capacity), phosphorus limitation (25 μM) and their combination in four faba bean varieties grown in greenhouse. The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

Leaf water potential

The treatments of water stress and/or P deficiency reduced free water in leaves which means the need of high pressure to extract it. Our results showed a significant ($P < 0.001$, Table 6) decrease of leaf water potential under all treatments and it was most pronounced under the combined deficit. All the four varieties were affected by the stresses to a varying extent. LO variety was the least affected with 87% of water potential decrease under the combined water and P limitation, versus 98% for AI variety relative to their corresponding controls. Under water and P limiting conditions, the leaf water potential of varieties LO and Ag were -5.84 and -7.64 MPa respectively (Figure 6) in contrast to RM that showed the lowest leaf water potential with -12.16 MPa.

Membrane stability

Electrolyte leakage and MDA have been studied in order to have more data on membrane stability. The statistical analysis of these two parameters revealed a significant effect of water deficit and P limitation when separated ($P < 0.001$, Table 6) but no significant difference on MDA was observed with the combined treatment. Electrolyte leakage and MDA of the four varieties changed in a similar pattern after plant exposure to stresses (Figure 7).

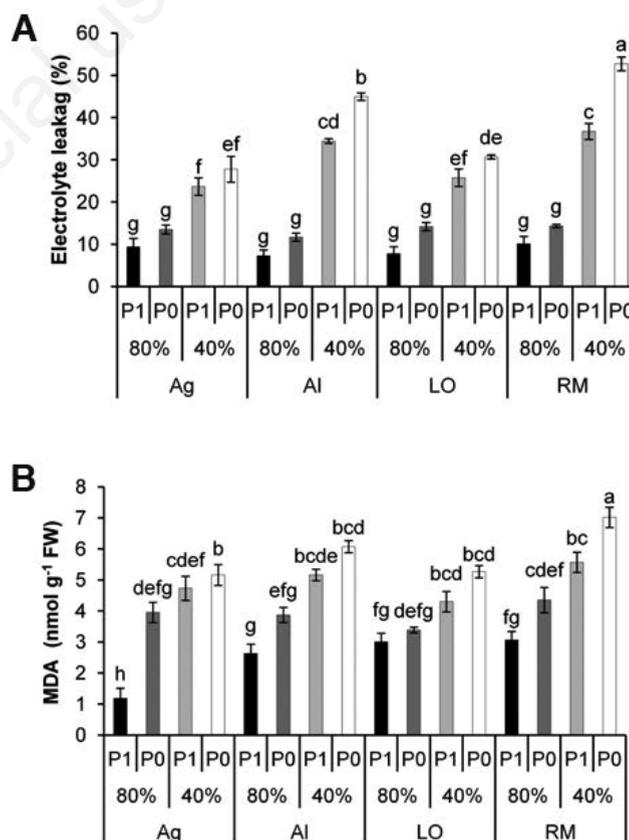


Figure 7. Electrolyte leakage (A) and malondialdehyde (B) accumulation as affected by water deficit, phosphorus deficiency and their combination in four faba bean varieties. The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

Water deficit induced an increase of these two parameters and the combination of both stresses caused greater increase in all of the tested varieties. Under water deficit combined with P deficiency the highest values of electrolyte leakage and MDA with 52.7% and 7 nmol g⁻¹ FW respectively, were obtained in RM variety. On the other hand, Ag variety presented the lowest electrolyte leakage value and MDA accumulation consisting of 27.8% and 5.16 nmol g⁻¹ FW respectively. These low values for Ag variety were significantly different from Al and RM varieties for electrolyte leakage but only from RM for MDA.

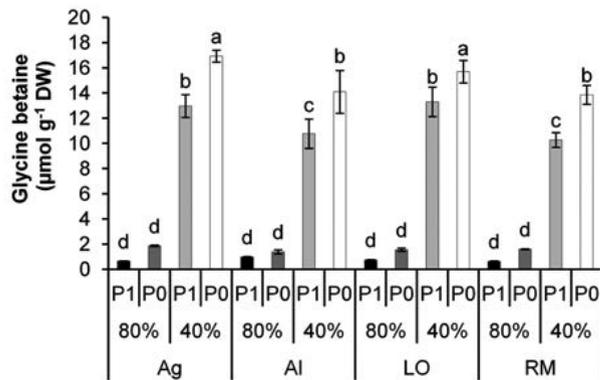


Figure 8. Glycine betaine accumulation under water deficit [40% field capacity (FC)], phosphorus deficiency (25 µM) and their combination (40% FC; 25 µM) in four faba bean varieties; Ag (Aguadulce), Al (Alfia), LO (Luz de Otono) and RM (Reina Mora). The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

Glycine betaine

The results presented in Figure 8 show the accumulation of glycine betaine (GB) in the four faba bean varieties. They revealed a high increase of glycine betaine accumulation under water deficit irrespective of P level. Therefore, GB accumulation responded more strongly to water deficit than to P deficiency. Nevertheless, the highest accumulation was obtained under water deficit combined with P shortage for all of the tested varieties and was reduced in the presence of phosphorus. Indeed, statistical analysis showed: i) a significant difference between the varieties ($P < 0.001$, Table 6) in response to water deficit for GB accumulation; ii) no significant differences were observed between the varieties in response to P limitation under adequate moisture; and so, iii) water deficit has a more marked effect than P deficiency. Our results showed that Ag and LO varieties accumulated the highest amounts of GB (16.92 and 15.69 µmol g⁻¹ DW respectively) while RM and Al varieties accumulated the lowest amounts (13.85 and 14.09 µmol g⁻¹ DW respectively) under the combination of both constraints. The presence of sufficient phosphorus under water deficit induced much reduced quantities of GB; 12.97 µmol g⁻¹ DW and 10.27 µmol g⁻¹ DW respectively for Ag and RM.

Plant nutrients concentrations

The statistical analysis revealed a significant ($P < 0.001$, Table 6) effect of water deficit and P deficiency on shoot, root and nodule P concentrations while no significant difference was observed under the combined water deficit and phosphorus deficiency on root P concentration between varieties. Limiting P had a greater effect on P concentration regardless of water regime.

LO variety showed the highest P concentration in all of the plant parts tested for the control treatment (0.44, 0.28 and 0.52 mg g⁻¹ DW respectively for shoot, root and nodule, Table 1). Ag variety P concentration showed a significant similarity to that of LO variety in the shoot. Under water deficit and P deficiency, P concentration in shoot decreased the least in the Ag variety (33.36%) and decreased the

Table 1. Shoot, root and nodule phosphorus concentrations under different phosphorus (P) treatments (P1; 125 µM versus P0; 25 µM), irrigation treatments [80% field capacity (FC) versus 40% FC] and their combination (25 µM, 40% FC) in four faba bean varieties grown in greenhouse conditions.

Varieties	Irrigation treatment	Phosphorus treatment	P-Shoot mg g ⁻¹ DW	P-Root mg g ⁻¹ DW	P-Nodule mg g ⁻¹ DW
Ag	80%	P1	0.41±0.04 ^{ab}	0.26±0.012 ^b	0.41±0.018 ^b
Ag	80%	P0	0.31±0.01 ^{de}	0.17±0.004 ^{fg}	0.28±0.029 ^{def}
Ag	40%	P1	0.34±0.01 ^{cd}	0.21±0.012 ^{cd}	0.38±0.005 ^b
Ag	40%	P0	0.27±0.01 ^{ef}	0.14±0.002 ^h	0.23±0.022 ^g
Al	80%	P1	0.37±0.04 ^{bc}	0.21±0.010 ^{cd}	0.34±0.031 ^c
Al	80%	P0	0.30±0.02 ^{de}	0.17±0.004 ^{ef}	0.26±0.029 ^{efg}
Al	40%	P1	0.32±0.01 ^{cde}	0.19±0.006 ^{def}	0.34±0.046 ^b
Al	40%	P0	0.21±0.02 ^{fg}	0.14±0.021 ^h	0.14±0.018 ⁱ
LO	80%	P1	0.44±0.04 ^a	0.28±0.024 ^a	0.52±0.013 ^a
LO	80%	P0	0.26±0.02 ^{ef}	0.20±0.004 ^{de}	0.25±0.013 ^{fg}
LO	40%	P1	0.28±0.03 ^{ef}	0.23±0.003 ^c	0.29±0.025 ^{de}
LO	40%	P0	0.26±0.06 ^{ef}	0.14±0.009 ^h	0.09±0.017 ^j
RM	80%	P1	0.32±0.04 ^{cde}	0.18±0.027 ^{def}	0.31±0.002 ^d
RM	80%	P0	0.22±0.02 ^{fg}	0.15±0.004 ^{gh}	0.19±0.013 ^h
RM	40%	P1	0.26±0.04 ^{ef}	0.18±0.013 ^{ef}	0.28±0.007 ^{def}
RM	40%	P0	0.19±0.03 ^g	0.13±0.005 ^h	0.10±0.007 ^j

Values are means of three replicates ± standard error. DW, dry weight; P, phosphorus; Ag, Aguadulce variety; Al, Alfia variety; LO, Luz d'Otono variety; RM, Reina Mora variety. ^{a-j} Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

most in Al variety (42.62%) relative to their respective controls. For the nodules, Ag variety had the highest P concentration (0.23 mg g⁻¹ DW) while LO and RM varieties showed the lowest ones with 0.09 and 0.10 mg g⁻¹ DW respectively. The statistical analysis revealed a significant individual effect of water deficit and P deficiency on shoot, root and nodule K⁺, Na⁺ and Ca²⁺ concentrations (P<0.001, Table 6). However, the combined effect of water deficit and P deficiency on the four varieties was statistically insignificant on Ca²⁺ concentration in shoot and K⁺ concentration in root. For all varieties (Ag, Al, LO and RM) the leaf K⁺ ions accumulated more than Na⁺

and Ca²⁺ in the controls and in the stressed plants (Table 2) while nodules accumulated Ca²⁺ in the highest concentrations for all treatments and varieties (Table 4). Na⁺ ions accumulated more in roots under both stresses (Table 3). Under the combined effect of water deficit and phosphorus limitation, Ag variety accumulated significantly more Ca²⁺ in its shoots, roots and nodules (18.75, 28.38, 39.93 mg g⁻¹ DW respectively), and Na⁺ in its nodules (24 mg g⁻¹ DW, Table 4). The lowest concentrations of Na⁺ and Ca²⁺ were observed in RM variety respectively in nodules (14.54 mg g⁻¹ DW) and shoots (9.29 mg g⁻¹ DW, Table 2) while Al variety had the low-

Table 2. Shoot K⁺, Na⁺ and Ca²⁺ concentrations of faba bean varieties under different phosphorus (P) treatments (P1; 125 µM and P0; 25 µM) and irrigation treatments [80% field capacity (FC) versus 40% FC].

Varieties	Irrigation treatment	Phosphorus treatment	Root nutrient concentration		
			K ⁺ mg g ⁻¹ DW	Na ⁺ mg g ⁻¹ DW	Ca ²⁺ mg g ⁻¹ DW
Ag	80%	P1	17.19±0.69 ^c	4.78±0.29 ^{de}	8.51±0.73 ^e
Ag	80%	P0	17.97±0.65 ^c	6.18±0.30 ^{abc}	11.49±0.60 ^d
Ag	40%	P1	21.81±0.59 ^{ab}	7.22±0.24 ^a	15.48±0.47 ^{bc}
Ag	40%	P0	23.22±0.28 ^a	7.26±0.54 ^a	18.75±0.59 ^a
Al	80%	P1	15.92±1.28 ^{cd}	4.35±0.21 ^{de}	9.12±0.72 ^e
Al	80%	P0	20.3±0.28 ^b	5.23±0.65 ^{cde}	9.62±0.88 ^e
Al	40%	P1	19.9±1.08 ^b	6.75±0.85 ^{ab}	11.75±0.82 ^d
Al	40%	P0	21.61±0.35 ^{ab}	7.21±0.65 ^a	12.85±0.66 ^{cd}
LO	80%	P1	17.01±1.97 ^c	4.77±0.33 ^{de}	8.69±0.52 ^e
LO	80%	P0	16.07±0.57 ^{cd}	6.52±0.22 ^{ab}	11.72±1.26 ^d
LO	40%	P1	20.6±0.52 ^b	6.55±0.09 ^{ab}	14.31±0.95 ^{bc}
LO	40%	P0	21.14±0.42 ^b	7.08±0.96 ^a	14.27±1.07 ^b
RM	80%	P1	14.6±1.72 ^d	3.93±0.31 ^e	7.47±0.53 ^e
RM	80%	P0	17.47±0.69 ^c	4.09±0.55 ^e	8.14±0.66 ^e
RM	40%	P1	19.81±0.33 ^b	4.95±0.37 ^{de}	8.42±0.52 ^e
RM	40%	P0	21.18±0.5 ^b	5.69±0.33 ^{bcd}	9.29±1.06 ^e

Values are means of three replicates ± standard error. DW, dry weight; P, phosphorus; Ag, Aguadulce variety; Al, Alfia variety; LO, Luz d'Otono variety; RM, Reina Mora variety. ^{a-e}Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

Table 3. Root K⁺, Na⁺ and Ca²⁺ concentrations of faba bean varieties under different phosphorus (P) treatments (P1; 125 µM and P0; 25 µM) and irrigation treatments [80% field capacity (FC) versus 40% FC].

Varieties	Irrigation treatment	Phosphorus treatment	Shoot nutrient concentration		
			K ⁺ mg g ⁻¹ DW	Na ⁺ mg g ⁻¹ DW	Ca ²⁺ mg g ⁻¹ DW
Ag	80%	P1	7.31±0.77 ^h	11.83±0.53 ^f	10.8±1.07 ^{gh}
Ag	80%	P0	8.72±0.56 ^{fg}	14.89±0.47 ^d	15.42±1.70 ^d
Ag	40%	P1	10.23±0.67 ^{bcde}	17.45±0.17 ^a	21.49±1.57 ^b
Ag	40%	P0	12.44±0.24 ^a	17.6±0.42 ^a	28.38±0.76 ^a
Al	80%	P1	8.99±0.57 ^{efg}	15.78±0.26 ^{bcd}	8.51±0.62 ⁱ
Al	80%	P0	9.75±0.45 ^{def}	16.71±0.65 ^{ab}	11.56±0.44 ^{fg}
Al	40%	P1	11.64±0.98 ^{ab}	15.74±0.04 ^{bcd}	12.23±0.21 ^{efg}
Al	40%	P0	11.24±0.8 ^{abc}	17.6±0.36 ^a	14.28±0.82 ^{de}
LO	80%	P1	8.38±0.66 ^g	10.57±0.50 ^g	10.4±0.86 ^{ghi}
LO	80%	P0	11.13±0.55 ^{abc}	14.84±0.1 ^d	11.45±0.32 ^{fg}
LO	40%	P1	11.61±0.41 ^{ab}	16.08±0.66 ^d	13.66±0.38 ^{def}
LO	40%	P0	12.19±0.93 ^a	17.6±0.36 ^{bc}	20.44±0.88 ^b
RM	80%	P1	8.9±0.4 ^{efg}	13.25±0.73 ^e	8.99±0.70 ^{hi}
RM	80%	P0	9.89±0.89 ^{cdef}	14.68±0.56 ^d	13.64±0.46 ^{def}
RM	40%	P1	10.95±0.56 ^{abcd}	15.43±0.07 ^{cd}	18.5±0.32 ^c
RM	40%	P0	11.96±0.28 ^a	16.1±0.48 ^{bc}	20.49±0.87 ^b

Values are means of three replicates ± standard error. DW, dry weight; P, phosphorus; Ag, Aguadulce variety; Al, Alfia variety; LO, Luz d'Otono variety; RM, Reina Mora variety. ^{a-hi}Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

est concentration of Ca^{2+} in roots ($14.18 \text{ mg g}^{-1} \text{ DW}$, Table 3) and nodules ($26.63 \text{ mg g}^{-1} \text{ DW}$). Under sufficient water and phosphorus nutrition, the significant variations between the varieties were observed in Na^+ concentration in roots and nodules. In roots, the highest concentration was observed in Al variety ($15.78 \text{ mg g}^{-1} \text{ DW}$) and the lowest one was observed in LO variety ($10.57 \text{ mg g}^{-1} \text{ DW}$). LO variety produced the highest concentration in nodules ($7.92 \text{ mg g}^{-1} \text{ DW}$) while RM variety had the lowest Na^+ concentration ($3.16 \text{ mg g}^{-1} \text{ DW}$).

Plant nitrogen concentration

Ag variety had the significantly highest concentration in shoot (4.65% , Figure 9A) under sufficient water and P treatment. For water deficit and phosphorus deficiency, they had a significant individual effect ($P < 0.001$, Table 6) on shoot and root N concentration. As Figure 9 shows, the combined effect of water deficit and P deficiency relative to the control, produced a significant reduction rate of 20% of N concentration in shoot of Ag variety and of 29.8%, 26.5 and 30% respectively in Al, LO and RM root N concentrations.

Correlation

All parameters studied were correlated significantly ($P < 0.01$, Table 5). Shoots represent the most important part of the plant for ensuring a good yield, because of their role as a source for nutrients and their translocation to the seeds. Nodules are also important because they ensure nitrogen fixation resulting from symbiosis with nitrogen fixing rhizobia. For these reasons we focused on shoot and nodule correlation to all the studied parameters. Shoot dry weight was more positively correlated to nodule dry weight ($r = 0.951$), stomatal conductance ($r = 0.824$), shoot and root nitrogen concentrations ($r = 0.623$; $r = 0.638$) and negatively correlated to electrolyte leakage ($r = -0.848$), MDA ($r = -0.891$), glycine betaine ($r = -0.839$). While, nodule dry weight was positively correlated to leaf area ($r = 0.819$), leaf water potential ($r = 0.808$), stomatal conductance ($r = 0.819$) and relative to root N concentration ($r = 0.523$)

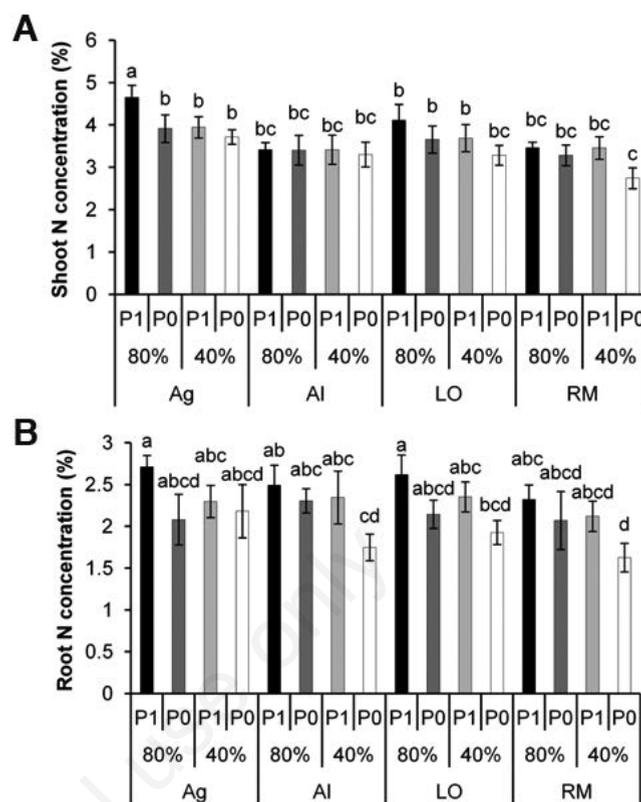


Figure 9. Shoot (A) and root (B) nitrogen concentration under different phosphorus treatments (125 μM versus 25 μM), irrigation treatments [80% field capacity (FC) versus 40% FC] and their combination (25 μM , 40% FC) in four faba bean varieties grown in greenhouse conditions. The values represent the means of three replications and the bars represent standard errors. Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

Table 4. Nodule K^+ , Na^+ and Ca^{2+} concentrations of faba bean varieties under different phosphorus (P) treatments (P1; 125 μM and P0; 25 μM) and irrigation treatments [80% field capacity (FC) versus 40% FC].

Varieties	Irrigation treatment	Phosphorus treatment	Nodule nutrient concentration		
			K^+ $\text{mg g}^{-1} \text{ DW}$	Na^+ $\text{mg g}^{-1} \text{ DW}$	Ca^{2+} $\text{mg g}^{-1} \text{ DW}$
Ag	80%	P1	9.93 ± 1.03^d	4.41 ± 0.15^j	12.81 ± 1.25^i
Ag	80%	P0	11.19 ± 1.96^d	8.92 ± 0.21^g	14.41 ± 0.49^{hi}
Ag	40%	P1	13.33 ± 1.45^{bc}	15.35 ± 1.52^c	33.53 ± 0.90^b
Ag	40%	P0	17.41 ± 1.14^a	24 ± 0.08^a	39.93 ± 0.07^a
Al	80%	P1	4.68 ± 0.34^e	6.98 ± 0.11^i	13.76 ± 0.13^{hi}
Al	80%	P0	9.65 ± 1.6^d	11.62 ± 0.10^d	13.82 ± 1.48^{hi}
Al	40%	P1	9.85 ± 0.78^d	12.05 ± 0.87^d	14.89 ± 2.22^g
Al	40%	P0	14.88 ± 2.87^b	16.79 ± 0.18^b	26.63 ± 1.20^d
LO	80%	P1	10.28 ± 1.72^d	7.92 ± 0.08^h	15.46 ± 0.06^{sh}
LO	80%	P0	11.79 ± 1.57^{cd}	10.29 ± 0.22^{ef}	17.21 ± 1.09^{efg}
LO	40%	P1	13.41 ± 1.45^{bc}	10.97 ± 0.02^{de}	19.42 ± 0.99^e
LO	40%	P0	16.85 ± 1.54^b	17.35 ± 0.56^b	27.40 ± 2.48^d
RM	80%	P1	5.56 ± 0.13^e	3.16 ± 0.23^k	15.87 ± 0.55^{sh}
RM	80%	P0	10.59 ± 0.27^d	9.51 ± 0.18^{fg}	17.25 ± 1.45^{efg}
RM	40%	P1	11.60 ± 0.63^{cd}	10.97 ± 0.57^{de}	18.87 ± 0.70^{ef}
RM	40%	P0	14.12 ± 0.85^b	14.54 ± 0.69^c	30.38 ± 2.17^c

Values are means of three replicates \pm standard error. DW, dry weight; P, phosphorus; Ag, Aguadulce variety; Al, Alfia variety; LO, Luz d'Otono variety; RM, Reina Mora variety. ^{a-j}Values with different letters are significantly different at 0.05 level of probability by Student-Newman-Keuls.

Table 5. Correlation coefficient of the studied parameters.

	RWC	Leaf_area	Shoot_DW	Root_DW	Nodule_DW
RWC	1	0.716**	0.768**	0.646**	0.743**
Stomatal conductance	0.728**	0.746**	0.824**	0.772**	0.819**
Leaf water potential	0.735**	0.651**	0.782**	0.773**	0.808**
Fv/Fm	0.806**	0.562**	0.727**	0.747**	0.726**
Electrolyte_leakage	-0.768**	-0.698**	-0.848**	-0.777**	-0.855**
MDA	-0.697**	-0.670**	-0.891**	-0.759**	-0.838**
Glycine betaine	-0.736**	-0.732**	-0.839**	-0.778**	-0.907**
N_Shoot	0.488**	0.485**	0.623**	0.330*	0.510**
N_Root	0.618**	0.474**	0.638**	0.538**	0.523**
P_Shoot	0.609**	0.615**	0.690**	0.537**	0.613**
P_Root	0.587**	0.657**	0.653**	0.418**	0.574**
P_Nod	0.617**	0.615**	0.609**	0.452**	0.525**
K_Shoot	-0.740**	-0.746**	-0.693**	-0.580**	-0.756**
K_Root	-0.711**	-0.653**	-0.807**	-0.667**	-0.818**
K_Nod	-0.623**	-0.489**	-0.574**	-0.676**	-0.607**
Ca_Shoot	-0.449**	-0.426**	-0.444**	-0.529**	-0.515**
Ca_Root	-0.517**	-0.569**	-0.624**	-0.517**	-0.656**
Ca_Nod	-0.540**	-0.537**	-0.681**	-0.579**	-0.710**
Na_Shoot	-0.564**	-0.423**	-0.505**	-0.651**	-0.563**
Na_Root	-0.640**	-0.813**	-0.685**	-0.378**	-0.685**
Na_Nod	-0.755**	-0.665**	-0.709**	-0.609**	-0.731**

RWC, relative water content; DW, dry weight; MDA, malondialdehyde. The values represent the correlation coefficient (n=48), **Very significant degree at 0.01 level of probability.

Table 6. Three-ways analyses of variance (ANOVA) of the variety (V), phosphorus treatment (P), irrigation treatment (I) and their interaction for the studied parameters.

	Dependent variables			Independent variables			
	V	P	I	V×P	V×I	P×I	V×P×I
Shoot_DW	122.982***	433.672***	4079.898***	22.836***	45.074***	48.030***	74.654***
Root_DW	10.088***	113.397***	238.855***	7.827**	23.586***	2.954ns	21.336***
Nodule_DW	149.67***	222.762***	17008.531***	0.584ns	215.141***	101.084***	2.818ns
Leaf_area	62.439***	411.178***	687.169***	29.853***	43.90***	192.728**	1.875ns
RWC	14.016**	69.366***	158.207***	1.177ns	1.094ns	0.761ns	2.849ns
Stomatal_conductance	38.646***	126.727***	1424.749***	6.094**	23.710***	28.241***	3.765*
Leaf_water_potential	32.12***	652.312***	3295.754***	47.974***	89.621***	168.026***	57.14***
Fv/Fm	4.11ns	42.446***	227.449***	4.311*	2.243ns	26.166***	4.492*
Electrolyte_leakage	23.325***	69.409***	1160.551***	2.462ns	43.597***	6.202*	3.464*
MDA	18.107**	85.019***	126.003***	9.785**	1.419ns	3.449ns	1.113ns
Glycine_betaine	18.030**	69.223***	5754.868***	0.775ns	18.948***	24.344***	0.352ns
N_Shoot	15.43**	15.663***	31.049***	1.178ns	2.866ns	0.010ns	1.399ns
N_Root	2.672 ns	41.021***	18.19**	0.092ns	0.485ns	0.033ns	2.663ns
P_Shoot	11.917**	139.317***	88.200***	0.098ns	1.452ns	7.695*	7.127**
P_Root	21.261***	278.361***	276.545***	10.12**	24.788***	0.048ns	1.237ns
P_Nodule	58.853***	26.29.349***	138.465***	48.101***	27.584***	44.587***	48.143***
K ⁺ _Shoot	5.487***	46.625***	247.359***	9.86**	3.688ns	1.347ns	4.615*
K ⁺ _Root	3.521ns	45.947***	279.051***	4.709*	4.58*	3.357ns	3.623ns
K ⁺ _Nodule	18.037**	133.91***	738.451***	3.751*	1.762ns	0.904ns	3.836*
Ca ²⁺ _Shoot	84.824***	68.786***	231.894***	9.810**	22.089***	0.06ns	2.514ns
Ca ²⁺ _Root	601.908***	163.356***	1191.046***	5.051**	72.444***	3.180ns	9.318**
Ca ²⁺ _Nodule	105.513***	376.381***	1069.535***	3.805*	119.253***	219.267***	6.259**
Na ⁺ _Shoot	89.609***	38.651***	57.515***	1.449ns	1.191ns	6.362*	3.459*
Na ⁺ _Root	106.567***	139.209***	346.99***	6.423**	41.146***	24.804***	10.722***
Na ⁺ _Nodule	73.564***	1354.923***	2075.034***	12.289***	136.254***	23.889***	35.509***

Numbers represent F values at 5% level. *Significant; **very significant; ***highly significant. NS, not significant; V, varieties; P, phosphorus treatment; I, irrigation treatment; DW, dry weight; RWC, relative water content; MDA, malondialdehyde.

and more negatively correlated to electrolyte leakage ($r = -0.855$) and MDA ($r = -0.838$), glycine betaine ($r = -0.907$). Thus, from evaluation across the majority of the parameters studied, Aguadulce variety was less affected by the combined effect of water deficit and P deficiency since it showed the highest shoot and nodule dry weights, the lowest decrease rate of stomatal conductance relative to the control, the highest leaf area and GB accumulation and the lowest electrolyte leakage and MDA under combined stresses in comparison to the other varieties. Hence, it could be considered as the most tolerant variety. On the other hand, Reina Mora variety, based on the same parameters, appeared to be the variety least tolerant to combined water and P limitation.

Discussion

Plants biomass and yield are the main parameters characterizing agricultural production. However, many environmental stresses such as water deficit and phosphorus limitation cause a series of changes in plants that adversely affect these desirable characters. Many reports have evaluated the individual effect of these abiotic stresses in various plant species including legumes. However, the need for studies on their combined effect lead us to study not only the individual effect of water deficit and P deficiency but also their combined limitation on faba bean defence and metabolism and then examine the performance of four different varieties under drought and/or P stresses. According to our results plant DW (shoot, root and nodule) as well as the leaf area were reduced under water deficit and P limitation in the four studied faba bean varieties, and the reductions were more pronounced under the combined stresses with a significant difference between the varieties. Indeed, the Ag variety was found tolerant compared to others, and this was reflected by low dry biomass reduction and a large foliar area under the combined stressors. According to Antolín *et al.* (2010) the reduction of plant dry biomass under stress condition is explained by stomatal closure in order to limit plant water loss by transpiration which leads to a reduction in CO₂ assimilation and as a result, a reduction in the translocation of assimilates to the nodules. This mechanism also limits the assimilation of CO₂ by the plant and results in a reduction of photosynthesis and which leads to growth inhibition. Applied stresses, water deficit or phosphorus limitation, reduced plant relative water content and leaf water potential. This was accompanied with a reduction in the stomatal conductance, and the reductions were more pronounced under the combined stressors in all studied varieties particularly in RM. Moreover, significant positive correlations were observed between leaf relative water content, leaf water potential, stomatal conductance and plant growth (plant biomass and leaves area) (Table 5). Similar results were reported by Mouradi *et al.* (2016) on alfalfa-rhizobia symbiosis under drought stress. Furthermore, Fujita *et al.* (2003) showed that phosphorus limitation depressed tomato plant biomass and this was accompanied with a reduction in stomatal conductance. The reduction of photosynthetic activity represented by maximum quantum yield of PS II was explained by the reduction of leaf area and plant water content, under the combined action of both stresses. This was previously mentioned by Li *et al.* (2006) and by Singh and Reddy (2014) who showed that leaf chlorophyll fluorescence responses to drought and phosphorus deficiency by reducing F_v/F_m . Indeed, the enhanced application of P improves root development resulting in increased water and nutrients uptake (Wang *et al.*, 2010) and then high leaf water potential, increased stomatal conductance and photosynthetic rate (Waraich *et al.*, 2011). These explained effects of P on

water status, nutrient uptake, stomatal conductance and photosynthetic rate highlight the mechanism that could be related to the response of plants to the shortage of this nutrient element and also to water deficit.

The high correlation of shoot DW to nodule DW ($r = 0.951$) shows the importance of the symbiosis for plant growth, as legume-rhizobia symbiosis is sensitive to both water and P limitation (Sulieman *et al.*, 2013; Nasr Esfahani *et al.*, 2014) and that 20% of plant total P is assigned to nodule for biological nitrogen fixation (Gunawardena *et al.*, 1992). Moreover, reduced phosphorus application from 125 μmol to 25 μmol of P caused a significant decrease of phosphorus assimilation and BNF, particularly when the phosphorus deficiency was combined with drought, reflected by a reduction in plant nitrogen concentration in the four tested faba bean varieties since our plants were grown depending on BNF. Under the combined stresses, the lowest values of both P and N clearly reflected the decrease in plant growth and this is related to the influence of low soil moisture on root growth, nutrient mobility in the soil and the plant's ability to get optimal amounts of nutrients (Waraich *et al.*, 2011). The high nodule P concentration could be explained by the fact that; i) nodules appear to take up P directly from nutrient solution (Al-Niemi *et al.*, 1998); ii) plants use nodules as a sink for P; and also that iii) nodules need high P concentration for symbiotic nitrogen fixation that is an energy demanding process (Israel, 1993). For nitrogen, the highest values were recorded in the shoots compared to roots; and that could be explained by the efficient translocation of the nitrogen fixed by the plant to the shoot. Bargaz *et al.* (2011) also reported the harmful effect of phosphorus deficiency on P and N concentrations and plant growth of *Phaseolus vulgaris* while Mouradi *et al.* (2016) presented this effect on *Medicago sativa* under drought. Therefore, according to these studies the combined stressors may affect P and N concentration and that has been correlated to plants growth, root growth and BNF.

Our results showed that membrane stability parameters, electrolyte leakage and accumulation of MDA, were higher under water stress but the damage was more pronounced under both deficit conditions. The highest accumulation under combined stresses were obtained in RM and according to Jiang and Huang (2001) and Kirnak *et al.* (2001) the high electrolyte leakage and MDA accumulation is explained by the reduction of relative water content resulting in leaf senescence and subsequently to the reduction of photosynthetic pigment function and chlorophyll. Under combined stresses, Ag variety presented the lowest electrolyte leakage and MDA accumulation reflecting low membrane damage which was correlated by Kabbadj *et al.* (2017) to a better growth performance under water deficit.

Under osmotic stress, including drought, plants adopt several mechanisms to adjust their internal osmotic potential. Osmoprotectant accumulation including glycine betaine is one of the main adaptive osmotic stress responses in plant (Kido *et al.*, 2019). Under 40% of substrate field capacity glycine betaine accumulates significantly in the leaves and this was more pronounced under the combined stresses particularly in Ag variety. This accumulation even at low concentrations protects photosynthesis and membrane integrity, stabilizes native structure of proteins and enzymes and detoxifies reactive oxygen radicals produced during stress (Murata *et al.*, 2007; Chen and Murata, 2011). The mechanisms of plants protection by GB under stress for better drought tolerance are related by Bohnert and Jensen (1996) to replacement of water by GB in biochemical reactions, thereby maintaining normal metabolism and then relatively normal growth as shown for Ag variety. Our data showed that drought stress alone or combined with phosphorus deficiency induced not only glycine betaine accumulation but also an increase in

the concentration of the three plant tissues in some inorganic compounds like Na^+ , K^+ and Ca^{2+} for which the highest amounts were observed in roots and nodules. Bargaz *et al.* (2015) stated that osmotic adjustment in legume is achieved by Na^+ sequestration particularly in nodules. Similarly, under drought stress Farissi *et al.* (2013) showed that the accumulation of some inorganic compounds particularly Na^+ and K^+ is associated drought tolerance in alfalfa. As these organic and inorganic compounds accumulate in response by the plant to the stress while the plant dry weight decreases, a significant negative correlation between these compounds with plant relative water content and DW was found (Table 5). This correlation is explained by the fact that inorganic compounds help plants to mediate stress by their important role in osmotic potential and osmotic adjustment (Silva *et al.*, 2010) through the activation of membrane ATPase enzyme required to drain back the nutrients lost during cell damage and controlling plant metabolism and development (Palta, 1990). In addition, Blum (2017) explained that these organic and inorganic solutes induce a reduction in water potential without complete turgor loss, and thus increase competitiveness and reflect an adaptive trait in support of crop yield under stress condition.

Our experiments, revealed a significant difference between the studied varieties' (Ag, Al, LO, RM) response to water deficit and phosphorus deficiency. This genotypic variation in response could be due to the physiological, anatomical (Gu *et al.*, 2003) and genetic differences and must be exploited to select the more adapted varieties. Available literature indicates variation between drought tolerant and susceptible varieties which may be due to the maintenance of tissue turgor, physiological traits, water uptake from soil and reduction in water loss through stomatal closure (Song *et al.*, 1995; Siddique *et al.*, 2000). Globally, our results showed that of the four varieties tested, Ag responded the best to the presence of phosphorus especially for nitrogen concentration even under water deficit. This is the effect of biological nitrogen fixation which is a high energy consuming process as the form of ATP requires large amount of phosphorus. In addition, the study of the parameter's correlation and the discussion of their mechanisms effects, suggested that the tolerance of Ag variety could be explained by the fact that P improved its root system resulting in the improvement of water absorption and nutrient assimilation (Phosphorus and Nitrogen) and the increase of photosynthetic rate and stomatal conductance. Glycine betaine as it accumulated more in Ag leaves than in RM leaves suggests that the cells may be trying to maintain an osmotic regulation, water potential gradient and tissue hydrostatic pressure under stress.

Conclusions

In conclusion, among the four faba bean varieties examined in this study, Ag variety was identified as the best performer in terms of plant biomass, leaf water status, membrane stability, osmolyte accumulation and nutrient uptake under water and P deficiency. These responses of faba bean plants to the combined action of both stresses compared to the individual stress factor, demonstrate the existence of considerable defence mechanisms among the varieties to cope with the combination of the two stresses.

The differences highlighted between *Vicia faba* varieties may be used to track down the genetic differences and the genes involved in the varieties' tolerance to water and phosphorus deficiency and which can also be used in breeding programs. These varieties could also be used for further assessment under field condition and for testing the alteration of both stresses on associated microorganism.

References

- Alghamdi S, Al-Shameri M, Migdadi H, Ammar M, El-Harty E, Khan M, Farooq M, 2015. Physiological and molecular characterization of faba bean (*Vicia faba* L.) genotypes for adaptation to drought stress. *J. Agron. Crop Sci.* 201:401-9.
- Al-Niemi TS, Kahn ML, McDermott TR, 1998. Phosphorus uptake by bean nodules. *Plant Soil.* 198:71-8.
- Antolín MC, Muro I, Sánchez-Díaz M, 2010. Application of sewage sludge improves growth, photosynthesis and antioxidant activities of nodulated alfalfa plants under drought conditions. *Environ. Exp. Bot.* 68:75-82.
- Argaw A, Mnalku A, 2017. Effectiveness of native Rhizobium on nodulation and yield of faba bean (*Vicia faba* L.) in Eastern Ethiopia. *Arch. Agron. Soil Sci.* 63:1390-403.
- Bargaz A, Drevon JJ, Oufdou K, Mandri B, Faghire M, Ghoulam C, 2011. Nodule phosphorus requirement and O₂ uptake in common bean genotypes under phosphorus deficiency. *Acta Agr. Scand. B-S. P.* 61:602-11.
- Bargaz A, Zaman-Allah M, Farissi M, Lazali M, Drevon JJ, Maougal RT, Georg C, 2015. Physiological and molecular aspects of tolerance to environmental constraints in grain and forage legumes. *Int. J. Mol. Sci.* 16:18976-9008.
- Blum A, 2017. Osmotic adjustment is a prime drought stress adaptive engine in support of plant production. *Plant. Cell. Environ.* 40:4-10.
- Bohnert HJ, Jensen RG, 1996. Strategies for engineering water-stress tolerance in plants. *Trends Biotechnol.* 14:89-97.
- Chen THH, Murata N, 2011. Glycinebetaine protects plants against abiotic stress: mechanisms and biotechnological applications. *Plant. Cell. Environ.* 34:1-20.
- Farissi M, Bouizgaren A, Faghire M, Bargaz A, Ghoulam C, 2013. Agrophysiological and biochemical properties associated with adaptation of *Medicago sativa* populations to water deficit. *Turk. J. Bot.* 37:1166-75.
- Food and Agriculture Organization of the United Nations, 2017. FAOSTAT Database. Available from: www.fao.org/faostat/ Accessed: 21 January 2018.
- Fujita K, Okada M, Lei K, Ito J, Ohkura K, Adu-Gyamfi JJ, Mohapatra PK, 2003. Effect of P deficiency on photoassimilate partitioning and rhythmic changes in fruit and stem diameter of tomato (*Lycopersicon esculentum*) during fruit growth. *J. Exp. Bot.* 54:2519-28.
- Ghoulam C, Foursy A, Fares K, 2002. Effects of salt stress on growth, inorganic ions and proline accumulation in relation to osmotic adjustment in five sugar beet cultivars. *Environ. Exp. Bot.* 47:39-50.
- Giri J, 2011. Glycinebetaine and abiotic stress tolerance in plants. *Plant Signal. Behav.* 6:1746-1751.
- Grieve CM, Grattan SR, 1983. Rapid assay for determination of water soluble quaternary ammonium compounds. *Plant Soil.* 70:303-7.
- Gu M, Rom CR, Robbins JA, 2003. Leaf gas exchange and stomatal characteristics of six birch taxa under difference irrigation regimes. *Ark. Agr. Exp. Sta. Res. Series.* 506:14-16.
- Gunawardena SFBN, Danso SKA, Zapata F, 1992. Phosphorus requirement and nitrogen accumulation by three mung bean (*Vigna radiata* L.) cultivars. *Plant Soil.* 147:267-74.
- Herridge DF, Peoples MB, Boddey RM, 2008. Global inputs of biological nitrogen fixation in agricultural systems. *Plant Soil.* 311:1-18.
- Hinsinger P, 2001. Bioavailability of soil inorganic P in the rhizosphere as affected by root-induced chemical changes: a review. *Plant Soil.* 237:173-95.

- Israel DW, 1993. Symbiotic dinitrogen fixation and host-plant growth during development of and recovery from phosphorus deficiency. *Physiol. Plantarum*. 88:294-300.
- Jaleel CA, Manivannan P, Wahid A, Farooq M, Somasundaram R, Panneerselvam R, 2009. Drought stress in plants: a review on morphological characteristics and pigments composition. *Int. J. Agric. Biol.* 11:100-5.
- Jemo M, Sulieman S, Bekkaoui F, Olomide OAK, Hashem A, Abd_Allah EF, Alqarawi AA, Tran L-SP, 2017. Comparative analysis of the combined effects of different water and phosphate levels on growth and biological nitrogen fixation of nine cowpea varieties. *Front. Plant. Sci.* 8:2111.
- Jensen ES, Peoples MB, Boddey RM, Gresshoff PM, Hauggaard-Nielsen H, Alves BJR, Morrison MJ, 2012. Legumes for mitigation of climate change and the provision of feedstock for biofuels and biorefineries. A review. *Agron. Sustain. Dev.* 32:329-64.
- Jiang Y, Huang B, 2001. Drought and heat stress injury to two cool-season turfgrasses in relation to antioxidant metabolism and lipid peroxidation. *Crop Sci.* 41:436-42.
- Jifon JL, Syvertsen JP, 2003. Moderate shade can increase net gas exchange and reduce photoinhibition in citrus leaves. *Tree Physiol.* 23:119-27.
- Kabbadj A, Makoudi B, Mouradi M, Pauly N, Frenedo P, Ghoulam C, 2017. Physiological and biochemical responses involved in water deficit tolerance of nitrogen-fixing *Vicia faba*. *PLoS One*. 12:e0190284.
- Kido ÉA, Ferreira-Neto JRC, da Silva MD, Santos VEP, da Silva Filho JLB, Benko-Iseppon AM, 2019. Osmoprotectant-related genes in plants under abiotic stress: expression dynamics, In silico genome mapping and biotechnology. In: M. Hossain, V. Kumar, D. Burritt, M. Fujita, P. Mäkelä (Eds.), *Osmoprotectant-mediated abiotic stress tolerance in plants*. Springer, Cham. 342:1-40.
- Kirkby EA, Johnston AE, 2008. Soil and fertilizer phosphorus in relation to crop nutrition. In: White P.J., Hammond J.P. (Eds.), *The ecophysiology of plant-phosphorus interactions*. *Plant Ecophysiology*, Vol. 7. Springer, Dordrecht, pp 177-223.
- Kimak H, Cengiz K, David H, Sinan G, 2001. A long-term experiment to study the role of mulches in physiology and macronutrition of strawberry grown under water stress. *Aust. J. Agricult. Res.* 52:937-43.
- Koivunen E, Partanen K, Perttilä S, Palander S, Tuunainen P, Valaja J, 2016. Digestibility and energy value of pea (*Pisum sativum* L.), faba bean (*Vicia faba* L.) and blue lupin (narrow-leaf) (*Lupinus angustifolius*) seeds in broilers. *Anim. Feed Sci. Technol.* 218:120-7.
- Kubure TE, Raghavaiah CV, Hamza I, 2016. Production potential of faba bean (*Vicia faba* L.) varieties in relation to plant densities and phosphorus nutrition on vertisols of Central Highlands of West Showa Zone, Ethiopia, East Africa. *Adv. Crop Sci. Tech.* 4:214.
- Li RH, Guo PG, Baumz M, Grando S, Ceccarelli S, 2006. Evaluation of chlorophyll content and fluorescence parameters as indicators of drought tolerance in barley. *Agr. Sci. China.* 5:751-7.
- Makoudi B, Kabbadj A, Mouradi M, Amenc L, Domergue O, Blair M, Drevon JJ, Ghoulam C, 2018. Phosphorus deficiency increases nodule phytase activity of faba bean-rhizobia symbiosis. *Acta Physiol. Plant.* 40:63.
- Mouradi M, Farissi M, Bouizgaren A, Makoudi B, Kabbadj A, Very AA, Sentenac H, Qaddoury A, Ghoulam C, 2016. Effects of water deficit on growth, nodulation and physiological and biochemical processes in *Medicago sativa*-rhizobia symbiotic association. *Arid Land Res. Manag.* 30:193-208.
- Mouradi M, Farissi M, Khadraji A, Makoudi B, Ghoulam C, 2018. Biochemical and antioxidant proprieties associated with the adaptation of faba bean (*Vicia faba* L.) - rhizobia symbiosis to phosphorus deficit. *J. Mater. Environ. Sci.* 9:1574-81.
- Multari S, Stewart D, Russell WR, 2015. Potential of Fava bean as future protein supply to partially replace meat intake in the human diet. *Compr. Rev. Food Sci. Food Saf.* 14:511-22.
- Murata N, Takahashi S, Nishiyama Y, Allakhverdiev SI, 2007. Photoinhibition of photosystem II under environmental stress. *B.B.A.-Bioenergetics.* 1767:414-21.
- Nasr Esfahani M, Sulieman S, Schulze J, Yamaguchi-Shinozaki K, Shinozaki K, Tran LS, 2014. Mechanisms of physiological adjustment of N₂ fixation in *Cicer arietinum* L. (chickpea) during early stages of water deficit: single or multi-factor controls. *Plant J.* 79:964-80.
- Neugschwandtner R, Ziegler K, Kriegner S, Wagentristl H, Kaul HP, 2015. Nitrogen yield and nitrogen fixation of winter faba beans. *Acta Agr. Scand. B-S. P.* 65:658-66.
- Palta JP, 1990. Stress interactions at the cellular and membrane levels. *Hort. Sci.* 25:1377.
- Razmjoo K, Heydarzadeh P, Sabzalian MR, 2008. Effect of salinity and drought stresses on growth parameters and essential oil content of *Matricaria chamomile*. *Int. J. Agric. Biol.* 10:451-4.
- Šavicka M, Skute N, 2010. Effects of high temperature on malondialdehyde content, superoxide production and growth changes in wheat seedlings (*Triticum aestivum* L.). *Ekologija.* 56:26-33.
- Shubhra JD, Goswami CL, Munjal R, 2004. Influence of phosphorus application on water relations, biochemical parameters and gum content in cluster bean under water deficit. *Biol. Plant.* 48:445-8.
- Siddique BMR, Hamid A, Islam MS, 2000. Drought stress effect on water relations of wheat. *Bot. Bull. Acad.* 41:35-9.
- Silva EN, Ferreira-Silva SL, Viégas RA, Silveira JAG, 2010. The role of organic and inorganic solutes in the osmotic adjustment of drought-stressed *Jatropha curcas* plants. *Environ. Exp. Bot.* 69:279-85.
- Singh AK, Bharati RC, Manibhushan NC, Pedpati A, 2013. An assessment of faba bean (*Vicia faba* L.) current status and future prospect. *African. J. Agric. Res.* 8:6634-41.
- Singh SK, Reddy VR, 2014. Combined effects of phosphorus nutrition and elevated carbon dioxide concentration on chlorophyll fluorescence, photosynthesis, and nutrient efficiency of cotton. *J. Plant Nutr. Soil Sc.* 177:892-902.
- Song FB, Dai JY, Gu WB, Li HY, 1995. Effect of water stress on leaf water status in maize. *J. Jilin. Agric. Univ.* 17:5-9. [in Chinese].
- Sulieman S, Ha C Van, Schulze J, Tran, LSP, 2013. Growth and nodulation of symbiotic *Medicago truncatula* at different levels of phosphorus availability. *J. Exp. Bot.* 64:2701-12.
- Wang W, Vinocur B, Altman A, 2003. Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. *Planta.* 218:1-14.
- Wang X, Yan X, Liao H, 2010. Genetic improvement for phosphorus efficiency in soybean: a radical approach. *Ann. Bot.* 106:215-22.
- Waraich EA, Ahmad R, Ashraf MY, 2011. Role of mineral nutrition in alleviation of drought stress in plants. *Austral. J. Crop Sci.* 5:764-77.
- Zhu JK, 2002. Salt and drought stress signal transduction in plants. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 53:247-73.
- Zlatev Z, Lidon FC, 2012. An overview on drought induced changes in plant growth, water relations and photosynthesis. *Emir. J. Food Agr.* 24:57.